DITERPENOIDS AND FLAVONOIDS FROM CISTUS PALINHAE

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Abstract—Cativic, ladenic, labdanolic, 8α -hydroxy-13(E)-labden-15-oic and 3-phenylpropionic acids were isolated from Cistus palinhae. In addition two new acids were characterized as 8α -methoxy-labd-15-oic and (5R, 8R, 9S, 10S)-2-oxo-3-cis-cleroden-15-oic. From the neutral fraction were isolated the known 8,15-labdanediol, 8(17)labden-15-ol, 6-oxo-7-labden-15-ol and 6β -hydroxy-8(17)-labden-15-ol and also identified were the hydroxy derivatives 8α -hydroxy-15-phenylpropionoxy-labdane, 8α -hydroxy-15-acetoxy-labdane, 8-labden-15-ol and 8-epi-15-labdanediol. The weak acid fraction gave jaranol (5,4'-dihydroxy-3,7-dimethoxyflavone), genkwanin (5,4'-dihydroxy-3,7-methoxyflavone), 3-methylkampferol and 3-betuligenol.

INTRODUCTION

Cistus palinhae, which is found only in the extreme south west of the Iberian peninsula (Cape St Vincent, Portugal), together with C. clusii and C. heterophylus are the only members of the Cistus genus which grow on chalky soil; the others grow on acid soils.

C. palinhae is morphologically very similar to C. ladaniferus, known as 'jara', which is to be found on acid soils in the south and west of Spain. The major component of both plants is labdanolic acid, but in C. palinhae, 6-oxocativic acid, abundant in C. ladaniferus [1], was not isolated.

RESULTS AND DISCUSSION

The acid fraction of a hexane extract of C. palinhae, which was soluble in sodium bicarbonate solution, was esterified and separated by dry chromatography of the methyl esters into compounds 1–6. In the ¹H NMR spectrum of 3 a three proton singlet of a methyl ether appeared at δ 3.03 and another singlet centered at 1.01 was due to a methyl geminal to the methoxyl group. Its mass spectrum (M⁺ at m/z 352, $C_{22}H_{40}O_3$) corresponded to the methyl ester of a bicyclic diterpene acid with a tertiary methoxyl group. The structure of methyl 8 α -methoxylabd-15-oate (3) was confirmed by synthesis from methyl labdanolate by treatment with methyl iodide and sodium hydride [2].

Compound 6 was a ketoester with an unsaturated α , β -carbonyl group (IR 1730, 1660 and 1620 cm⁻¹, UV 234 nm, ϵ = 14500). The ¹H NMR spectrum showed signals of five methyl groups (two Me-C, two Me-CH and one Me-C= at δ 1.82) and a wide singlet of an olefinic hydrogen at 5.52.

The mass spectrum of **6** (M⁺ at m/z 334, C₂₁H₃₄O₃) corresponded to a bicyclic diterpene with a C₇ side chain (m/z at 129, C₇H₁₃O₂) and it was identical to the mass spectrum of the methyl ester of oxopopulifolic acid [3]. The fragment at m/z 205 corresponds [4] to a decalin with four methyl groups, a carbonyl group and a double bond. By double irradiation of a ¹H NMR signal centered at

 δ 5.52 the doublet at 1.84 was changed into a singlet and vice versa, confirming the presence of the -CO-CH =CMe grouping, which together with the existence of only two methyl singlets indicated a clerodane or cistane skeleton with the carbonyl group at C-2 and the double bond at C-3.

The CD curve of 6 in hexane showed maxima at 342 nm $(\Delta \varepsilon = +0.94)$, 229 nm $(\Delta \varepsilon = -4.45)$ and 206 nm $(\Delta \varepsilon = +6.31)$. Applying the reversed octant law of Snatzke and that of the helicity of transoid enones, the curve is only compatible with possibilities I, II and III.

The signal of the ¹H NMR spectrum [C₆D₆, Eu (dpm₃)] corresponding to the hydrogen on C-10 (dd, $J_{AX+BX} = 14$ Hz) excludes possibility I. The stereochemistry 5R, 8R, 9S, 10S for II was determined [2] by applying the McConnell and Robertson ratio [5]. However, this is still to be confirmed by X-ray diffraction.

The neutral fraction of a hexane extract of *Cistus palinhae* was resolved by dry chromatography. From the less polar fraction, a hydroxyester 7 with an aromatic system was obtained (IR 1610, 1500 cm⁻¹; ¹H NMR δ 7.09, 5H, br s) whose molecular ion in the mass spectrum was at m/z 442, which corresponds to the formula $C_{29}H_{46}O_3$. Alkaline hydrolysis of 7 yielded 8 [6] and β -phenylpropionic acid identified as a methyl ester by comparison with an authentic sample.

Compounds 7 and 9 were isolated from fraction II (see Experimental). Compound 9 was a hydroxyester with an acetoxyl group, and had the formula $C_{22}H_{40}O_3$ (M⁺ at m/z 352). Compound 9 was obtained by acetylation of 8. The more polar fraction IV contained 8 (8,15-labdane-diol).

Saponification of the neutral part yielded the unsaponifiable part which was fractionated by silica gel chromatography. Compounds 10–14 and sitosterol were isolated. Compound 10 showed signals in its 1H NMR spectrum of the following groupings: $-CH_2-CH_2OH$ (δ 3.53, 2H, t, J = 6 Hz), Me-C= (1.51, s, 3H) and four methyl groups (three Me-C and one Me-CH). Treatment of 8 with formic acid gave 15 which by alkaline hydrolysis yielded 10.

Ⅲ 5R.10S (10C.)

Compounds 11, 12 and 13 were identified by comparison with authentic samples [7]. Compound 14 of $[\alpha]_D + 6.3^\circ$ was a dihydroxyderivative with signals for the following groupings in its ¹H NMR spectrum: $-\text{CH}_2\text{OH}$ (δ 3.60, t, 2H, J = 6 Hz), Me-C-OH (δ 1.11, s, 3H), Me-C (0.97, 0.87 and 0.83, each s, 3H) and Me-CH (0.89, d, 3H, J = 6.5 Hz).

I 55.10 R (10C,)

The mass spectrum showed the [M]⁺ at m/z 310, corresponding to $C_{20}H_{38}O_2$. In the monoacetyl derivative of 16 the ¹H NMR signal of the methyl geminal to the hydroxyl group remained centered at δ 1.11. Compound

14 was given the structure of 8β ,15-labdanediol, thus accounting for the deshielding of the C-20 protons (δ 0.83 in 8 and 0.97 in 14 and 16). The spectroscopic properties of 14 were identical to those of 8α ,15-ent-labdanediol, obtained by reduction of the methyl ester of the 8α -ent-labda-15-oic acid, isolated from Trachylobium verrucosum [8].

III 5*5*.10*5* ("c₀)

Three flavonoids and one phenol were isolated from the weak acid fraction, soluble in sodium hydroxide and identified by comparison with authentic samples [9]. The flavonoids were jaranol (5,4'-dihydroxy-3,7-dimethoxy-

flavone), genkwanin (5,4'-dihydroxy-7-methoxyflavone) and 3-methylkampferol (5,7,4'-trihydroxy-3-methoxyflavone) and the phenol was betuligenol or (-)1(4-hydroxyphenyl)-butan-3-ol [10]. The primary components of the acidic part were β -phenylpropionic and labdanolic acid. No $D(-)\beta$ -hydroxy acids were detected; these are very important components of Cistus ladaniferus [11].

EXPERIMENTAL

Mps are uncorr. and were determined on a Kofler hot stage apparatus. UV spectra were recorded in EtOH. ¹H NMR spectra were recorded in CCl₄ using TMS as internal standard. Analytical TLC was performed on silica gel, prep. TLC was on silica gel PF₂₅₄₊₃₆₆ and CC was on silica gel 60. The CD curves were carried out on a Yobin Yvon Dicrograph III dicrograph using hexane as the solvent. The mass spectra were carried out with a source temperature of 180° and ionization energy of 70 eV.

Extraction and isolation. The aerial parts of C. palinhae (10 kg) collected at Cape St Vincent (Portugal) were dried and extracted with n-hexane in a Soxhlet for 24 hr giving 840 g of extract. This was dewaxed with MeOH (16%) and later extracted with 6% NaHCO₃ (26.2%) and 4% NaOH (0.5%), a neutral fraction remaining which represented 68.7% of the original extract. Treatment of 22.0 g of the bicarbonate-soluble acid part with CH₂N₂, yielded 21.8 g of a mixture of methyl esters, which were dry-chromatographed (770 g of silica gel, eluted with n-hexane-Et₂O, 1:1) giving four fractions.

Treatment of fraction I (2.2 g) with a satd methanolic soln of urea, gave 1.2 g of methyl esters soluble in MeOH, which were resolved by CC on AgNO₃-silica gel (2:8), yielding pure samples of 1 and 2. From fraction II (1.9 g) the methyl ester 3 was isolated by CC, followed by prep. TLC (C_6H_6 -CHCl₃, 9:1). Crystallization of fraction III (7.4 g) from hexane yielded pure methyl labdanolate (4). CC of the residual part (in AgNO₃-silica gel (2:8) gave 6, 5 and 4. Compounds 5 and 6 were isolated by prep. TLC (C_6H_6 -CHCl₃-Et₂O, 1:3:1).

(5R,8R,9S,10S)-Methyl-2-oxo-3-cis-cleroden-15-oate (6). Colourless oil. $[\alpha]_{D}^{22} - 3^{\circ}$ (CHCl₃; c 0.56). UV λ_{\max}^{EtOH} nm: 234 (£12 800). CD $\lambda_{\max}^{n-bexane}$ nm: 342 ($\Delta \epsilon$ + 0.94), 329 ($\Delta \epsilon$ - 4.45), 206 ($\Delta \epsilon$ + 6.31). IR ν_{\max}^{film} cm⁻¹: 1730, 1660, 1610, 1425, 1370, 1260, 1220, 1140, 1000, 840. ¹H NMR: δ 5.52 (1H, s, H-3), 3.67 (3H, s), 1.84 (3H, d, J = 1.5 Hz), 1.12 (3H, s), 0.98 (3H, s). EIMS 70 eV, m/z (rel. int.): 334 [M] + (6), 205 (39), 125 (38), 121 (95), 109 (100), 95 (90), 69 (40). The neutral fraction (8.2 g) was chromatographed on a dry column eluting with C₆H₆-Et₂O (7:3) to yield four fractions: I (38.5%); II (17.5%); III (8.0%) and IV (22.3%).

The NaOH soluble fraction was saturated with a stream of CO₂ yielding a precipitate which was separated by filtration and from which three flavonoids were isolated by CC on silica gel–NaOAc (92:8): jaranol, genkwanin and 3-methylkampferol. The aq. soln, after separating the flavonoids was acidified and extracted with Et₂O and (–)betuligenol was isolated by silica gel

8-Hydroxy-15-β-phenylpropionoxy-labdane (7). Fraction I was composed exclusively of 7. Colourless oil. $[\alpha]_{D}^{22} - 5.8^{\circ}$ (CHCl₃; c 5.70). IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 3510, 1740, 1610, 1500, 1160, 1080, 940, 910, 785, 750, 700. ¹H NMR: δ7.09 (5H, s), 4.03 (2H, t, J = 6 Hz), 2.69 (4H, m), 1.07 (3H, s), 0.85 (3H, s), 0.77 (3H, s), 0.89 (3H, d, J = 6.5 Hz). EIMS 70 eV, m/z (rel. int.): 442 [M]⁺ (3), 424 (2), 409 (2), 191 (7), 177 (13), 150 (63), 125 (34), 105 (63), 98 (100), 91 (63), 83 (55), 69 (95), 55 (46). Alkaline hydrolysis of 7 (200 mg, 10 ml NaOH–MeOH 10%) yielded 8 and β-phenylpropionic acid (the methyl ester identical to an authentic sample).

8-Hydroxy-15-acetoxy-labdane (9). Compound 9 was isolated

from fraction II by prep. TLC eluting with *n*-hexane–EtOAc. Colourless oil. $[\alpha]_{D}^{22} - 6.5^{\circ}$ (CHCl₃; *c* 2.60). IR $v_{\text{max}}^{\text{fin}}$ cm⁻¹: 3480, 1740, 1250, 1085, 1040, 940, 910. ¹H NMR: δ 4.02 (2H, t, J = 6 Hz), 1.96 (3H, s), 1.07 (3H, s), 0.91 (3H, d, J = 6.5 Hz), 0.86 (3H, s), 0.78 (6H, s). EIMS 70 eV, m/z (rel. int.): 352 [M] + (2), 329 (3), 324 (3), 177 (26), 164 (26), 150 (19), 137 (22), 125 (57), 109 (55), 95 (69), 83 (100), 60 (96), 61 (81), 55 (43).

8,15-Labdanediol (8). By crystallization in C_6H_6 , 8 was separated from fraction IV, mp 82–83°. $[\alpha]_D^{22}$ – 5.15° (CHCl₃; c 1.00). IR v_{\max}^{fin} cm⁻¹: 3350, 1130, 1080, 1050, 940, 910. ¹H NMR: δ 3.55 (2H, t, J = 6 Hz), 1.08 (3H, s), 0.90 (3H, d, J = 6 Hz), 0.87 (3H, s), 0.78 (6H, s), EIMS 70 eV, m/z (rel. int.): 310 [M] ⁺ (3), 292 (3), 277 (3), 177 (13), 157 (12), 137 (13), 123 (25), 109 (37), 95 (48), 83 (57), 81 (62), 60 (100), 55 (47).

A soln of 86 g of the neutral part in 200 ml NaOH-MeOH (10%) was left to stand overnight at room temp. The MeOH was evaporated, H_2O and HCl were added and the mixture was extracted with Et_2O . The ethereal soln was washed with 6% NaHCO₃ and 4% NaOH. The neutral part (61.2%) was chromatographed on silica gel, with C_6H_6 - Et_2O (9:1). A fraction (12.8%) was eluted which by later silica gel-AgNO₃ (8:2) CC and prep. TLC yielded 10, 11 and sitosterol. Compounds 12 and 13 were separated from the first fraction by CC on silica gel-AgNO₃ (8:2) eluted with C_6H_6 - Et_2O (1:1) (5.4%). Following this, elution with the same mixture gave a material (3.7%) which by crystallization in n-hexane gave 14. The remainder (76.9%) of the neutral fraction was 8.

8-Labden-15-ol (10). Colourless oil. $[\alpha]_{D}^{2D} + 31^{\circ}$ (CHCl₃; c 1.30). IR v_{\max}^{flm} cm⁻¹: 3330, 1470, 1380, 1060, 1020, 890. ¹H NMR: δ 3.53 (2H, t, J = 6 Hz), 1.51 (3H, s), 0.91 (3H, s), 0.89 (3H, d, d) = 6.5 Hz), 0.87 (3H, s), 0.72 (3H, s).

Treatment of 8,15-labdanediol (8) with HCO₂H. Compound 8 (111 mg) was treated with 1 ml HCO₂H at room temp. for 10 min, adding Et₂O dropwise to facilitate the dissolution of the product which appeared suspended in the soln. Et₂O was added, the soln was washed with 6% NaHCO₃ and H₂O. CC of the reaction product (110 mg) gave 15 (83 mg) and 10 (26 mg).

15-Formyloxy-8-labdene (15). Colourless oil. $[\alpha]_{D}^{22} + 40.5^{\circ}$ (CHCl₃; c 2.55). IR $v_{\text{max}}^{\text{flm}}$ cm⁻¹: 1740, 1715, 1460, 1360. ¹H NMR: δ 7.82 (1H, s), 4.10 (2H, t, J = 6 Hz), 1.50 (3H, s), 0.93 (3H, d, J = 6 Hz), 0.92 (3H, s), 0.89 (3H, s), 0.85 (3H, s). Alkaline hydrolysis of 15 yielded 10.

8(17)-Labden-15-ol (11). Colourless oil. $[\alpha]_D^{22} + 25.2^{\circ}$ (CHCl₃; c 1.40). IR $v_{\text{max}}^{\text{fim}}$ cm⁻¹: 3350, 3050, 1650, 1470, 1215, 1070, 895. ¹H NMR: δ 4.75 (1H, s br), 4.45 (1H, s br), 3.56 (2H, t, J = 6 Hz) 0.89 (3H, d, J = 6.5 Hz), 0.88 (3H, s), 0.81 (3H, s), 0.67 (3H, s). EIMS 70 eV m/z (rel. int.): 292 $[M]^+$ (8), 277 (10), 191 (9), 177 (36), 137 (61), 123 (39), 109 (41), 95 (87), 81 (100), 69 (57), 55 (61).

6-Oxo-7-labden-15-ol (12). Colourless oil. [α] $_D^2$ + 12° (CHCl₃; c 1.20). IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 3410, 1670, 1240, 1050, 1030, 970, 880. ¹H NMR: δ 5.62 (1H, s), 3.56 (2H, t, J = 6 Hz), 1.89 (3H, s), 1.10 (3H, s), 1.07 (3H, s), 0.91 (3H, d, J = 6.5 Hz), 0.81 (3H, s).

6β-Hydroxy-8(17)-labden-15-ol (13). Colourless oil. $[\alpha]_{D}^{22}$ + 23° (CHCl₃; c 1.30), IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 3380, 3080, 1650, 1120, 1040, 890, 860. ¹H NMR: δ4.93 (1H, s br), 4.73 (1H, s br), 4.31 (1H, m), 3.57 (2H, t, J = 6 Hz), 2.28 (2H, d, J = 3.5 Hz). EIMS 70 eV, m/z (rel. int.): 290 [M – 18] ⁺ (5), 275 (5), 153 (43), 135 (18), 109 (88), 95 (68), 81 (84), 69 (100), 55 (65).

8-Epi-15-labdanediol (14). Mp 137–140°. [α] $_{2}^{2}$ 2 + 6.3° (CHCl $_{3}$; c 0.97). IR ν $_{max}^{KBr}$ cm $^{-1}$: 3360, 1190, 1140, 1060, 1015, 910. 1 H NMR: δ 3.60 (2H, t, J = 6 Hz), 1.11 (3H, s), 0.97 (3H, s), 0.89 (3H, d, J = 6.5 Hz). 0.87 (3H, s), 0.83 (3H, s). EIMS 70 eV, m/z (rel. int.): 310 [M] $^{+}$ (3), 292 (4), 278 (5), 195 (13), 178 (26), 157 (24), 137 (15), 123 (35), 109 (60), 95 (52), 83 (66), 81 (66), 69 (100), 55 (63), 43 (76).

Compound 14 (20 mg) in 0.6 ml C₅H₅N was treated with 1 ml

of Ac₂O and left overnight to yield 20 mg 16. Colourless oil. $[\alpha]_D^{22} + 6.1^\circ$ (CHCl₃; c 0.97). IR $v_{\text{max}}^{\text{film}}$ cm⁻¹: 3530, 1745, 1250, 1140, 1060, 1040, 910. ¹H NMR: \dot{o} 4.07 (2H, t, J = 6 Hz), 2.02 (3H, s), 1.11 (3H, s), 0.97 (3H, s), 0.89 (3H, d, J = 6.5 Hz), 0.87 (3H, s), 0.82 (3H, s).

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